## Reactivation of *VaSTS1* expression in transgenic Arabidopsis thaliana plants by retransformation with 2b from Cucumber mosaic virus, isolate NK

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Since transgene silencing in genetically transformed plants is a serious limitation for a wide application of genetic engineering techniques, studying mechanisms ensuring stability of transgene expression is vital. Molecular mechanism underlying transgene silencing is RNA interference (RNAi). RNAi protects plant cells from the expression of viral and foreign DNA. Small interfering RNAs (siRNAs) are key components in RNAi. The siRNA-protein complexes inhibit transgene expression at the post-transcriptional and transcriptional levels by degrading target mRNA transcripts and establishing DNA methylation within transgene nucleotide sequences, respectively. Multiple investigations concerning viral suppressors of gene silencing revealed that 2b protein from Cucumovirus (CMV) effectively represses assembly and targeting of the RNA-induced silencing complex. Current study presents unique data on using the 2b gene from CMV-NK isolate for transgene silencing reduction in A. thaliana plants earlier transformed with VaSTS1. In our study, two VaSTS1-transgnenic lines were retransformed with 2b and derived plants were analyzed. Our data demonstrated that A. thaliana plants with decreased expression of VaSTS1 transgene increased transgene expression in up to 3.0-fold upon retransformation with 2b from CMV NK. Interestingly, the more pronounced effect of 2b retransformation regarding increase in transgene expression was shown for nptII used as selective marker for transformants selection upon transformation with VaSTS1 of A. thaliana plants. The nptII gene expression increased in more than 10.0-fold in lines retransformed with 2b being compared to initial plants transformed with VaSTS1. Moreover, the decrease in the level of VaSTS1 expression in transgenic A. thaliana plants was associated with enhancement of the cytosine DNA methylation level within VaSTS1 sequence. The mentioned fact implies that VaSTS1 expression was repressed at transcriptional level and our data demonstrates that 2b from CMV NK can reactivate a silenced transgene.

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